

What drives DNA folding into G-quadruplex structures and their recognition by ligands?

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Outline

Introduction

- DNA conformations
- Human telomeric (ht) DNA fragment Tel22 = AGGG(TTAGGG)₃
- G-quadruplex ligands

Method

- Global thermodynamic analysis of experimental unfolding and binding data

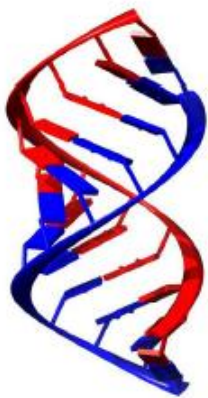
Results

- Driving forces of quadruplex folding and ligand binding

Conclusions

- Meaningful findings

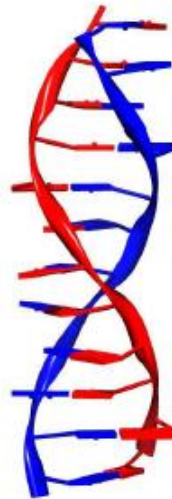
DNA conformations



A-DNA



B-DNA



Z-DNA

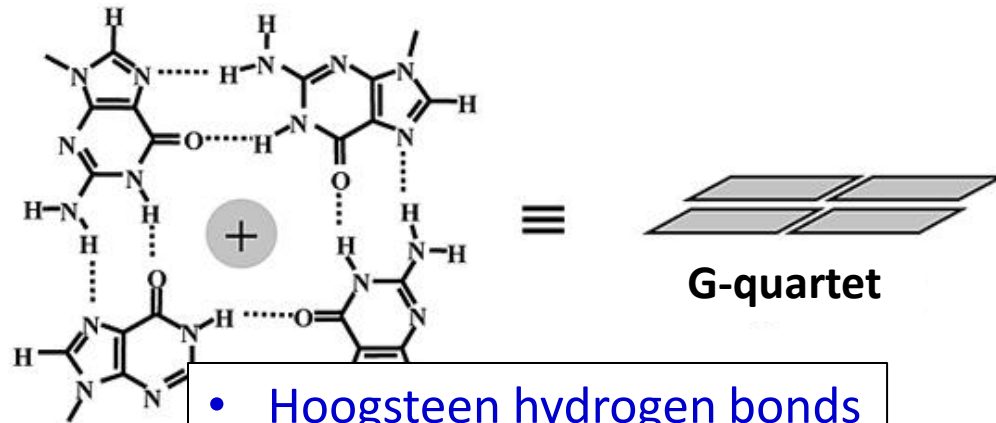


triplex

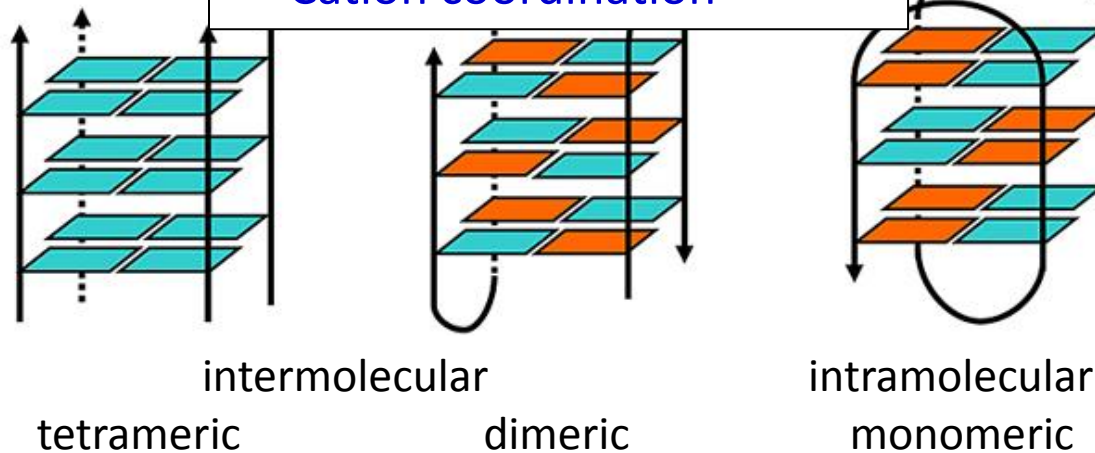


quadruplex

G-quadruplexes



- Hoogsteen hydrogen bonds
- Stacking interactions
- Cation coordination

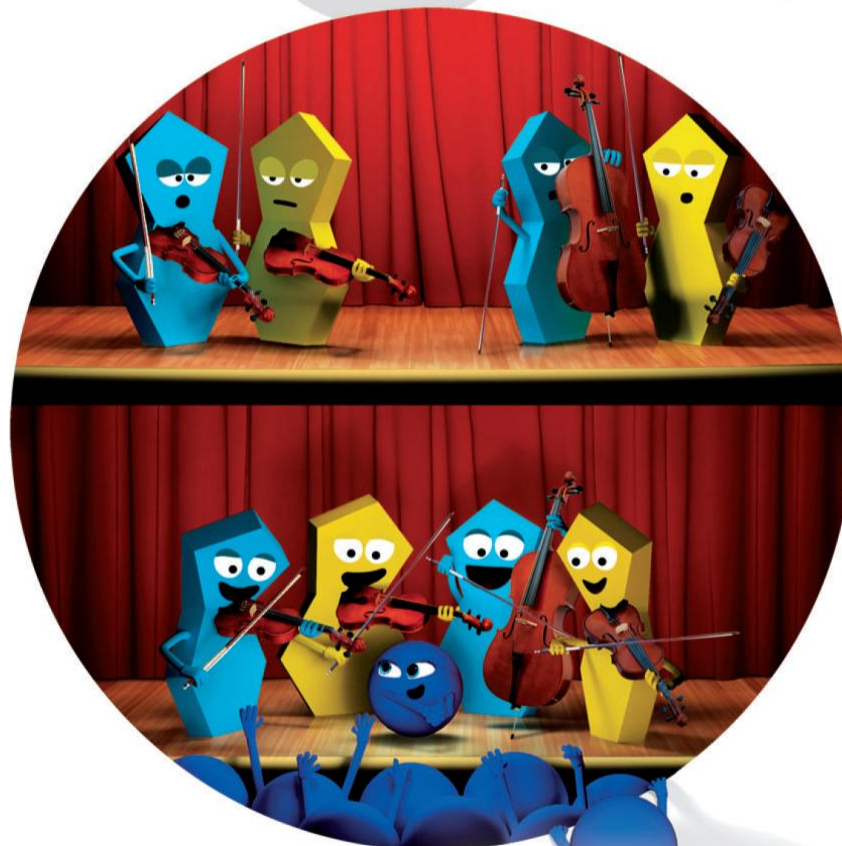


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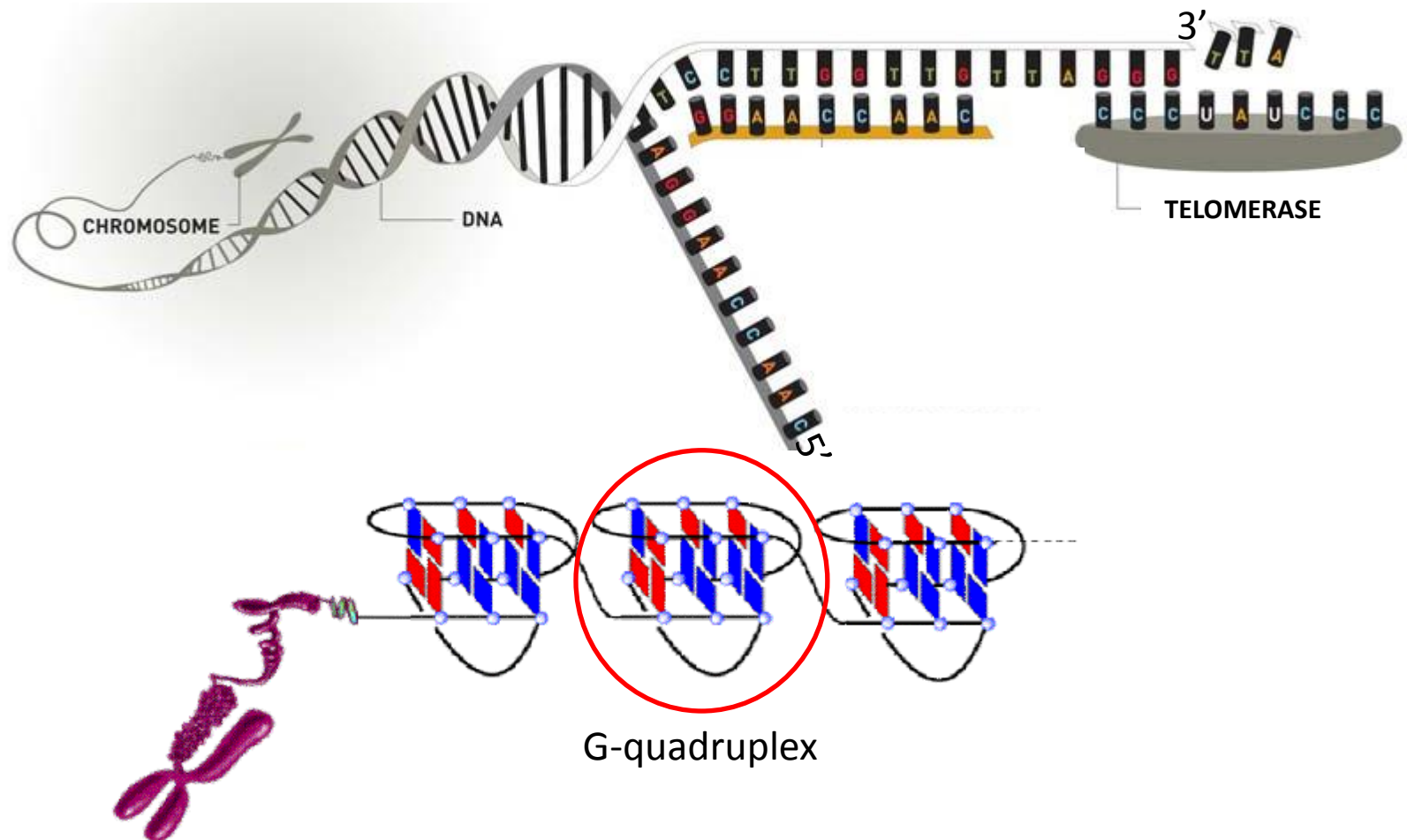
Guanine-rich DNA sequences can self-assemble into specific pre-organized DNA structures, where two by two guanine residues shown as “duets” are involved in GG-N1-carbonyl symmetric base pairs. In their Communication (DOI: 10.1002/anie.201400531), J. Lah, J. Plavec, and co-workers demonstrate the transition of these “duets” into guanine-quartets, presented as a “string quartet” when interacting with cations such as potassium ions. This rearrangement consequently leads to formation of G-quadruplex structures.

WILEY-VCH

Human telomeric DNA

$(TTAGGG)_N$

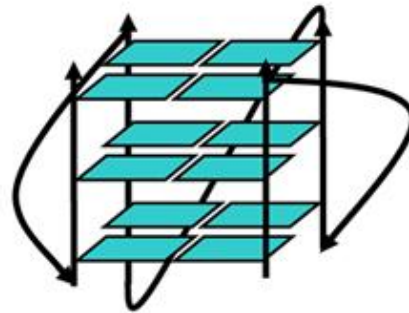
3'-overhang; 100-200 bases



Model system: Tel22 = AGGG(TTAGGG)₃



antiparallel
solution (Na⁺)



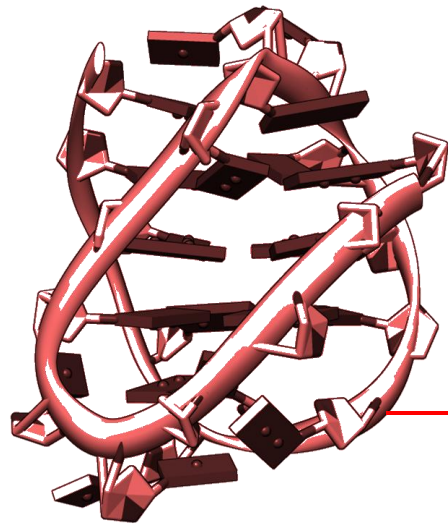
parallel
crystal (K⁺)
solution (K⁺, n.a. solvents, PEG)



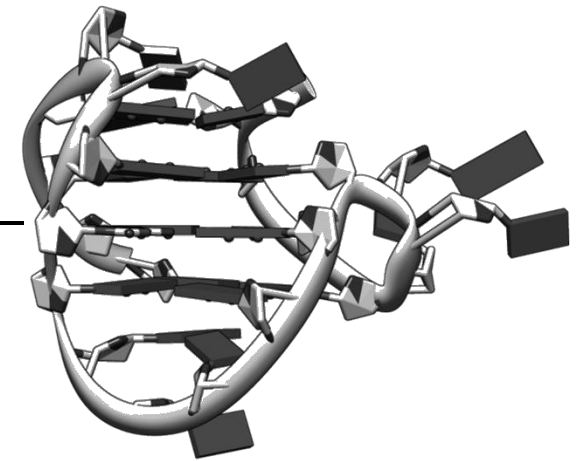
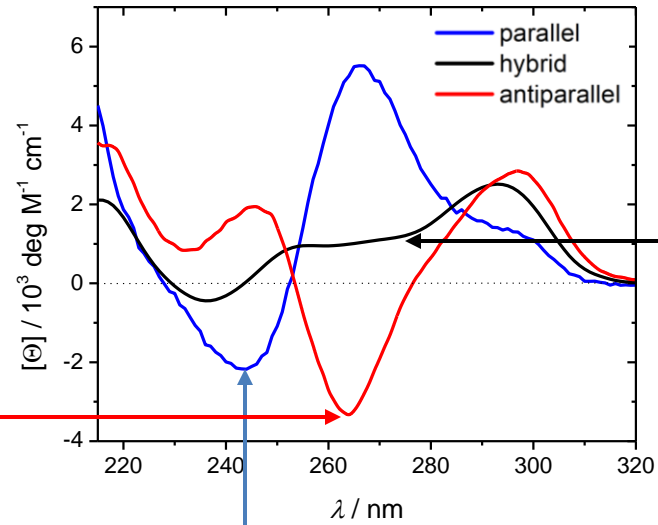
hybrid
solution (K⁺)

G.N. Parkinson et al. *Nature* 2002, **417**, 876; A. Ambrus et al. *Nucleic Acids Res.* 2006, **34**, 2723.
A.T. Phan et al. *Nucleic Acids Res.* 2007, **35**, 6517; R.D. Gray et al. *Biochemistry* 2010, **49**, 179.
K.N. Luu et al. *J. Am. Chem. Soc.* 2006, **128**, 9963; Y. Wang et al. *Structure* 1993, **1**, 263....

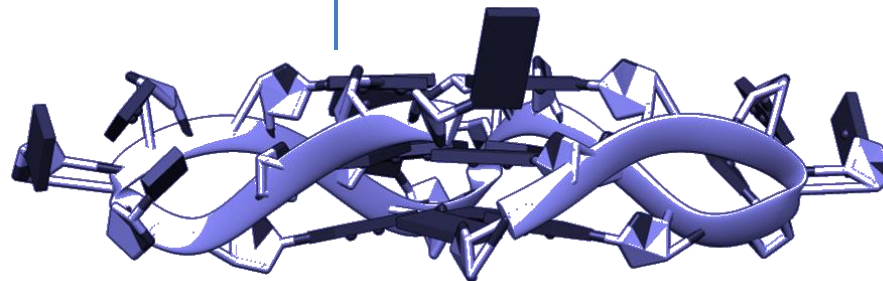
Structural features



antiparallel (A_{Na^+})

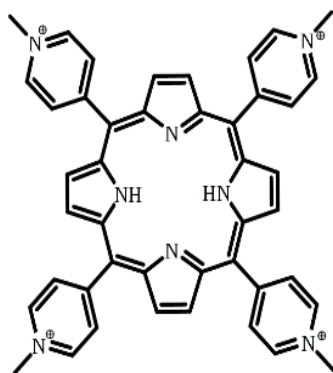
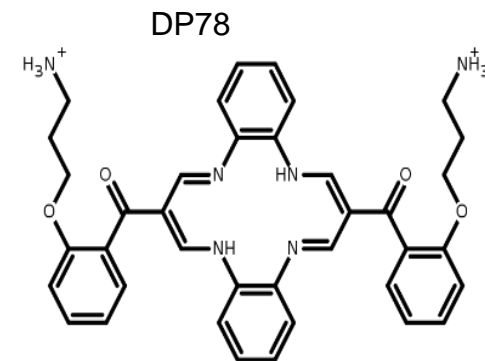
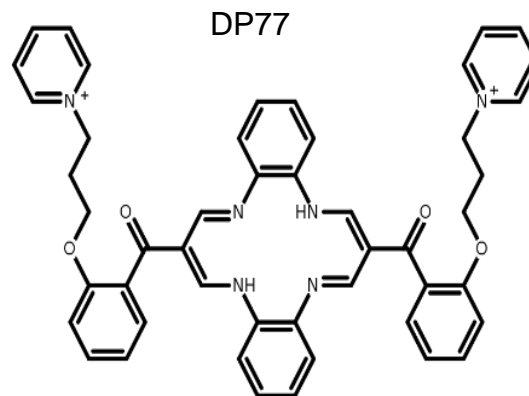
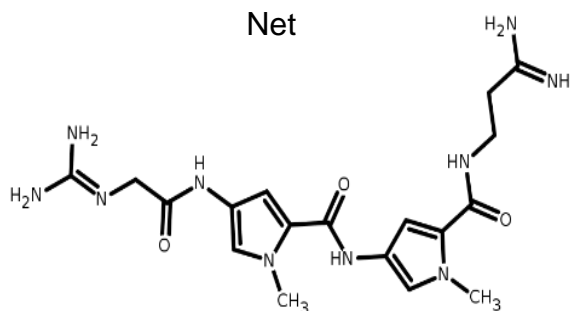


hybrid (H_{K^+})

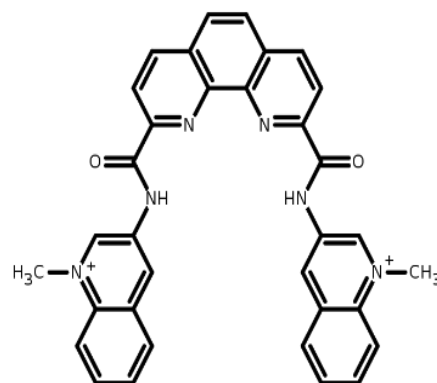


parallel (P_{K^+})

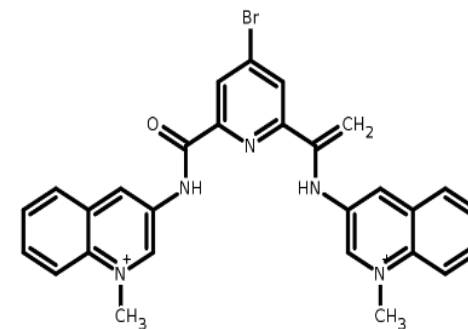
Ligands



TMPyP4



Phen-DC3



360A-Br

$$K_{\text{quadruplex}} / K_{\text{duplex}} > 50$$

Energetic basis of quadruplex folding and ligand binding

Why thermodynamic driving forces of quadruplex folding and ligand binding remain poorly understood?

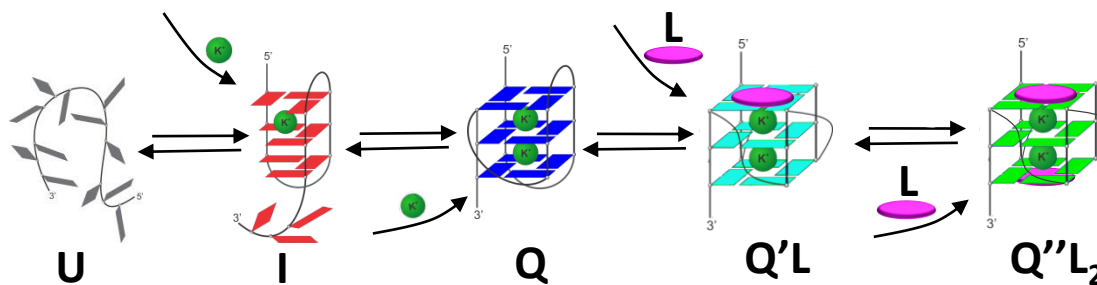
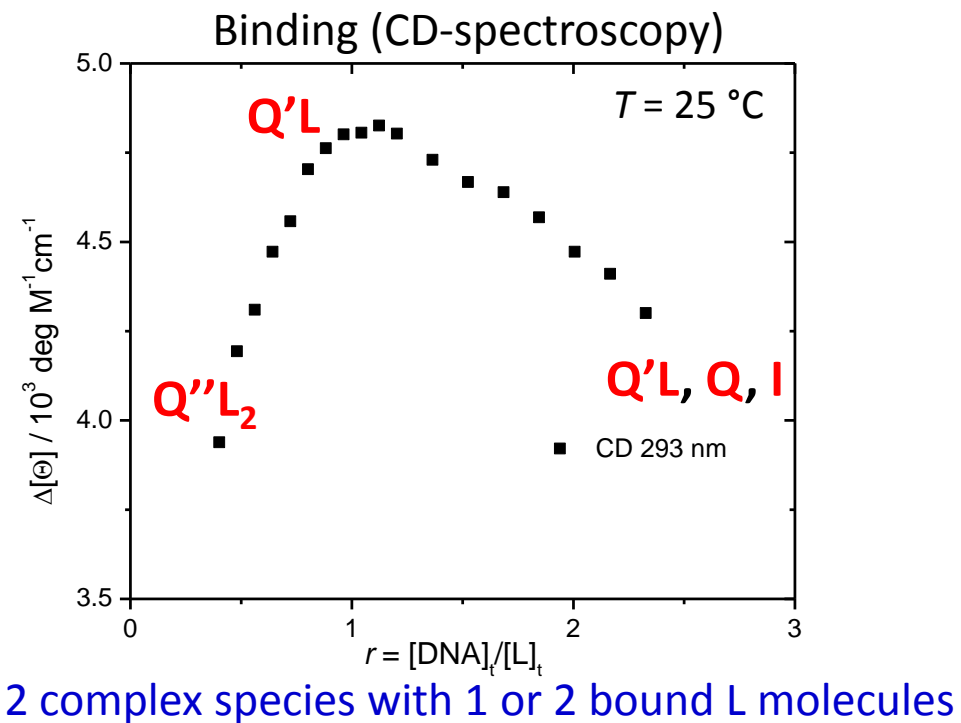
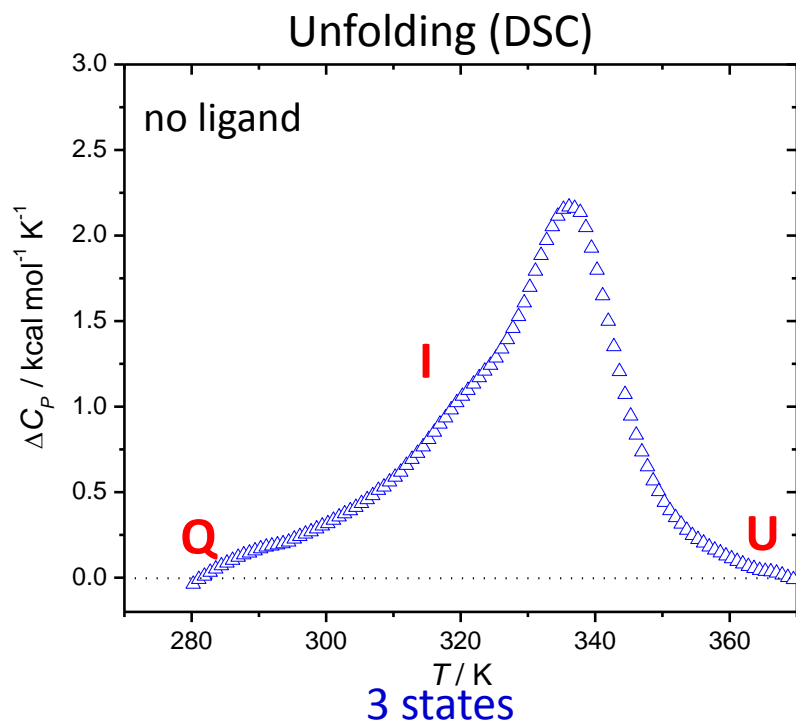
- Quadruplex folding is intrinsically complex: **Mechanism?**
- Hydration is a major determinant of DNA stability and conformation: $\Delta C_{p,i}$?
- Thermodynamic studies of ligand binding: **Folding intermediates?**
- “The role of “hydrophobic” forces in DNA folding in general is in fact well known?”
(Referee of our manuscript in 2014)

Energetic basis of quadruplex folding and ligand binding

Questions:

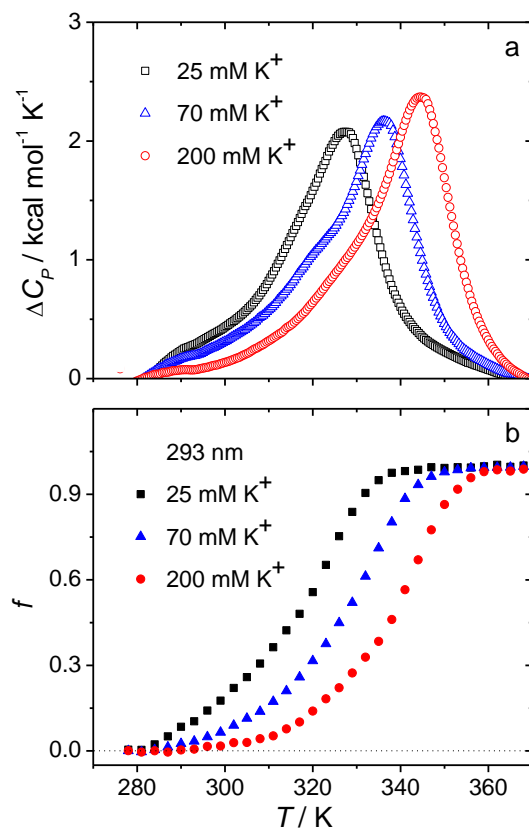
- What is the mechanism of folding and binding?
- What drives folding and binding events?
- Are predictions of thermodynamic analysis consistent with structural features?

Mechanism (Tel22, ligand = Phen-DC3, K⁺ ions)?

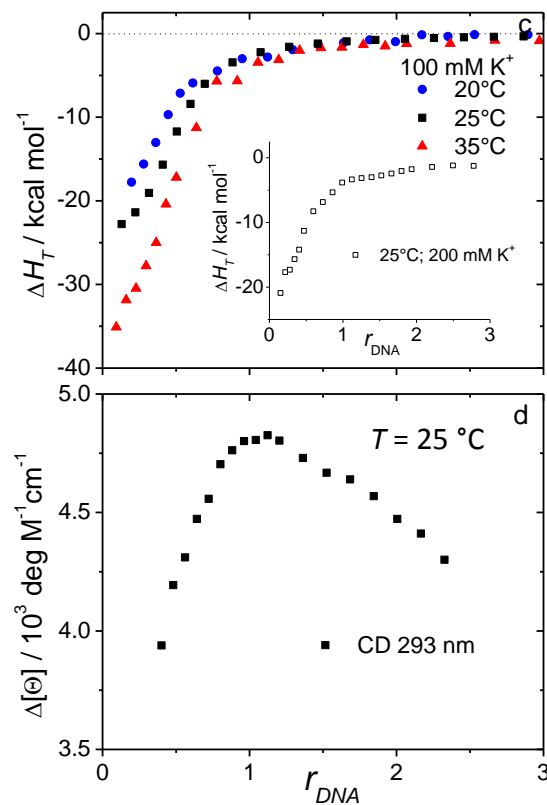


Calorimetry and spectroscopy

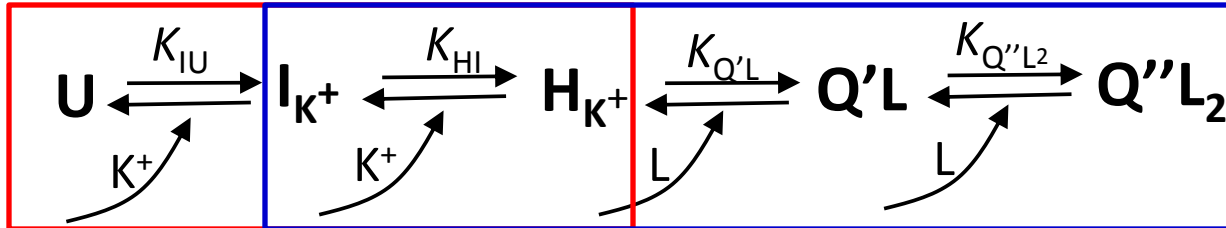
Melting (DSC, CD)



Titrations (ITC, CD)



Modeling folding and binding data



$$\text{DSC: } \Delta C_p = (\partial \alpha_I / \partial T) \Delta H_{FI(T)}^\circ + (\partial \alpha_U / \partial T) \Delta H_{FU(T)}^\circ$$

melting

$$\text{CD: } f = \frac{[\theta] - [\theta]_F}{[\theta]_I - [\theta]_F} = \alpha_I f_{FI} + \alpha_{II}$$

How the fractions (α_i) depend on temperature, DNA, salt, and ligand concentrations?

titrations

$$\left(\left(\frac{\partial n_I}{\partial n_{\text{DNA}}} \right)_{n_{L,\text{tot}}} - \left(\frac{\partial n_I}{\partial n_{\text{DNA}}} \right)_{n_{L=0}} \right) \Delta H_{QI(T)}^\circ$$

$$\text{CD: } \Delta[\theta] = \Delta[\theta]_{QI} \alpha_I + \Delta[\theta]_{Q'L} \alpha_{Q'L} + (\Delta[\theta]_{Q'L} + \Delta[\theta]_{Q''L_2}) \alpha_{Q''L_2}$$

Model functions are related to concentrations of species (fractions = α_i) at any temperature, DNA, salt, and ligand concentrations!

Thermodynamics of folding and binding

Adjustable parameters:

$$\Delta G_{ij(T_0)}^\circ = \Delta(\text{Gibbs free energy})$$

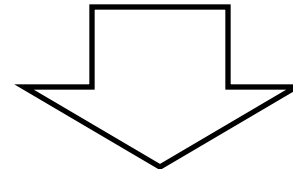
$$\Delta H_{ij(T_0)}^\circ = \Delta(\text{enthalpy})$$

$$\Delta C_{P,ij}^\circ = \Delta(\text{heat capacity})$$

$$n_{ij} = \Delta(\text{no. exchanged cations})$$

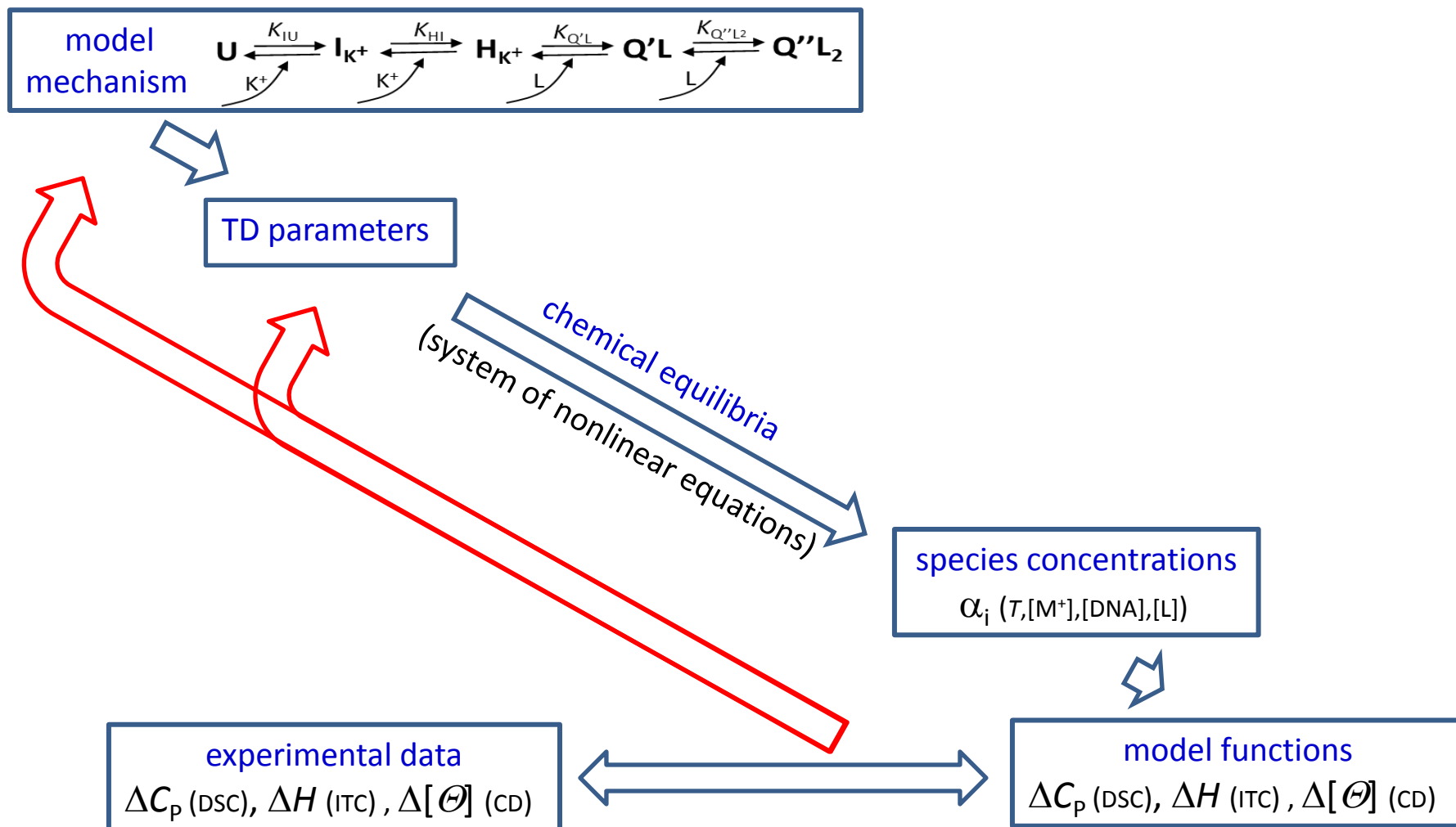
$$\Delta G_{ij(T,K^+)}^\circ = \Delta G_{ij(T_0)}^\circ T/T_0 + \Delta H_{ij(T_0)}^\circ [1 - T/T_0] + \Delta C_{P,ij}^\circ [T - T_0 - T \ln(T/T_0)] + n_{ij} RT \ln[K^+]$$

$$K_{ij(T,K^+)} = \exp(-\Delta G_{ij(T,K^+)}^\circ / RT) \Rightarrow \alpha_{i(T,K^+)} \Rightarrow \frac{\partial \alpha_i}{\partial T}, \frac{\partial \alpha_i}{\partial r}$$

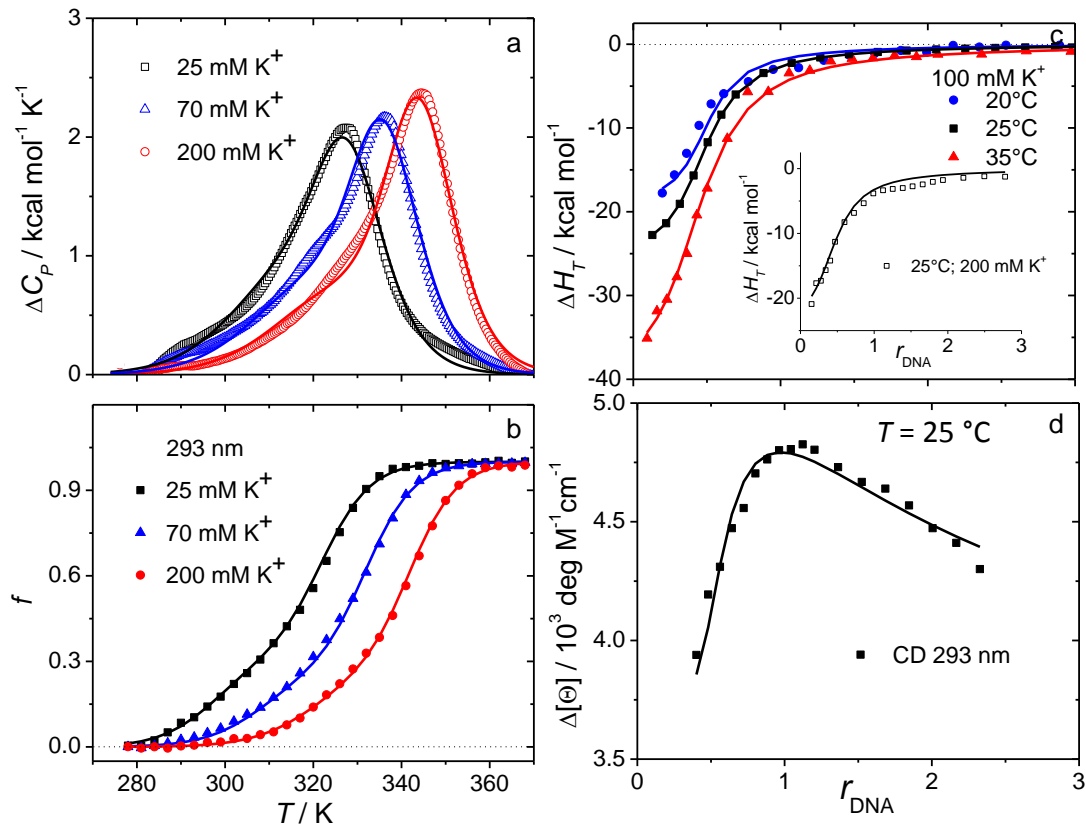
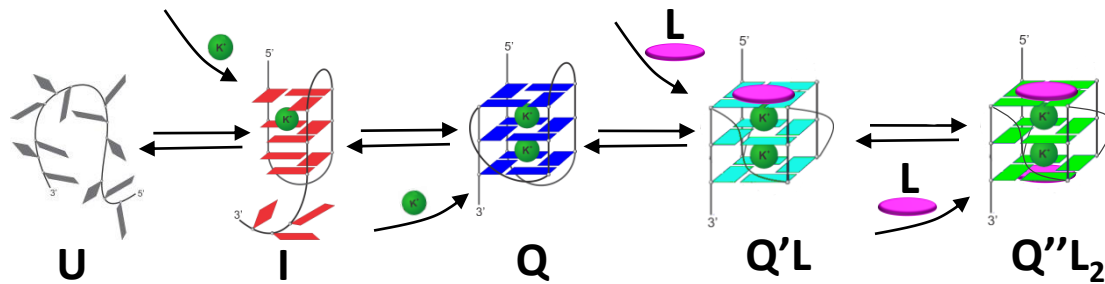


Model functions are completely defined at any temperature, DNA, salt, and ligand concentrations!

Global thermodynamics analysis



Global thermodynamics analysis



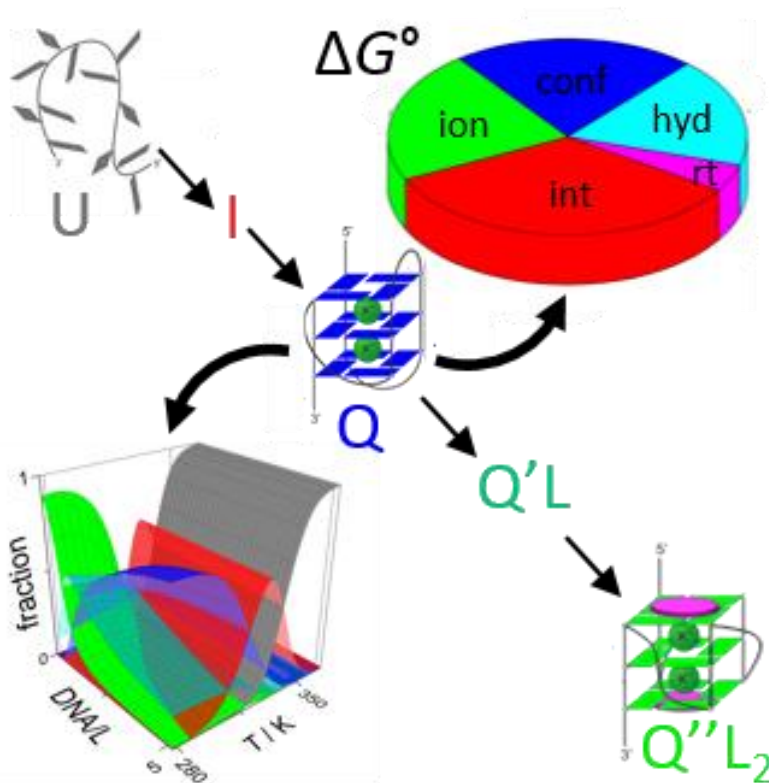
Advantages of global fitting

I. Drobnak et al. (2010). *J. Phys. Chem. B* **114**, 8713.

- More experimental points per fitted parameter
- Greater variety of data analyzed simultaneously
- More accurate parameter values
- **More rigorous test of model appropriateness**

Angewandte Chemie, 2014, 53, 4881; *ChemPhysChem*, 2014, 15, 1827.
J. Am. Chem. Soc., 2013, 135, 1288; *J. Biol. Chem.*, 2012, 287, 8613.
J. Am. Chem. Soc., 2012, 134, 9657; *Nucleic Acids Res.*, 2011, 39, 1933.
J. Agric. Food Chem., 2011, 59, 727; *J. Phys. Chem. B*, 2010, 114, 8713.
J. Phys. Chem. B, 2010, 114, 4313; *J. Biol. Chem.*, 2010, 285, 5606.
J. Biol. Chem., 2009, 284, 20002; *J. Mol. Biol.*, 2009, 392, 63.
J. Am. Chem. Soc., 2008, 130, 14161; *Nucleic Acids Res.*, 2008, 36, 897.
Biophys. J. 2015, in press...

Thermodynamics analysis ($\Delta G^\circ = \Delta H^\circ - T\Delta S^\circ$)



$$\Delta G^\circ = \Delta G^\circ_{\text{ion}} + \Delta G^\circ_{\text{hyd}} + \Delta G^\circ_{\text{int}} + \Delta G^\circ_{\text{rt}} + \Delta G^\circ_{\text{conf}}$$

$\Delta G^\circ_{\text{ion}}$... ion desolvation/solvation
(Y.A. Marcus, *Biophys. Chem.*, 1994, **51**, 111)

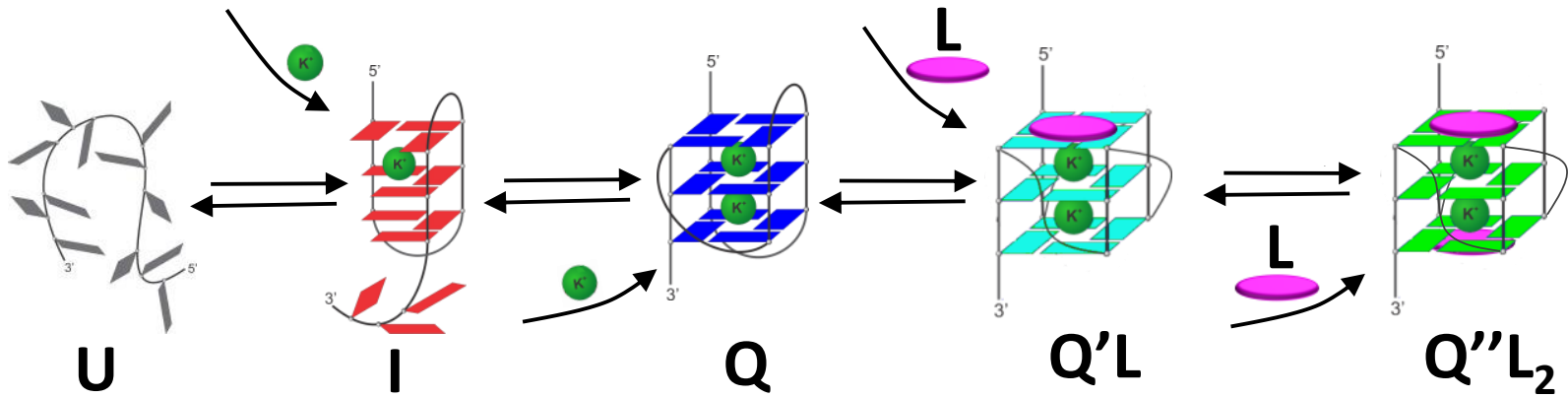
$\Delta G^\circ_{\text{hyd}}$... DNA and ligand desolvation/solvation
($\Delta G^\circ_{\text{hyd}} \approx \Delta C_p 80(\pm 10)\text{K}$ at 25 °C)

$\Delta G^\circ_{\text{int}}$... stacking, H-bonding, Coulombic interactions, cation coordination ($\Delta G^\circ_{\text{int}} \approx \Delta H^\circ - \Delta H^\circ_{\text{ion}}$)

$\Delta G^\circ_{\text{rt}}$... rotational and translational freedom of ligand and DNA (R.S. Spolar, M.T. Record, *Science*, 1994, **263**, 777)

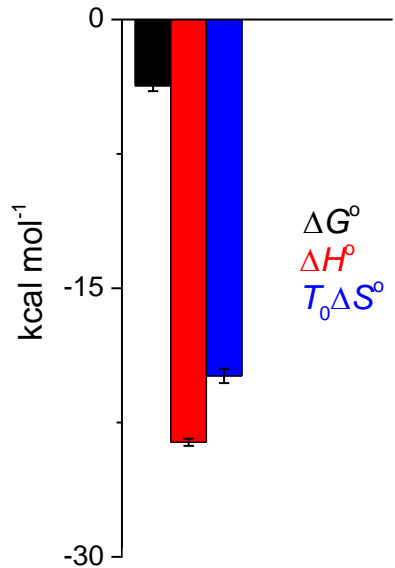
$\Delta G^\circ_{\text{conf}}$... conformation
($\Delta G^\circ_{\text{conf}} = \Delta G^\circ - \Delta G^\circ_{\text{solv}} - \Delta G^\circ_{\text{int}} - \Delta G^\circ_{\text{rt}}$)

Thermodynamic driving forces

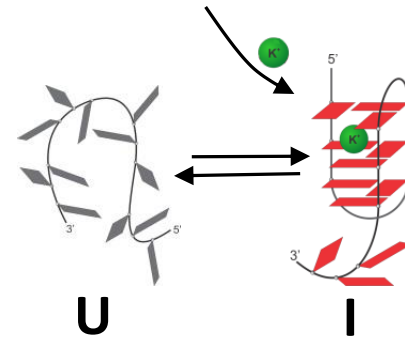
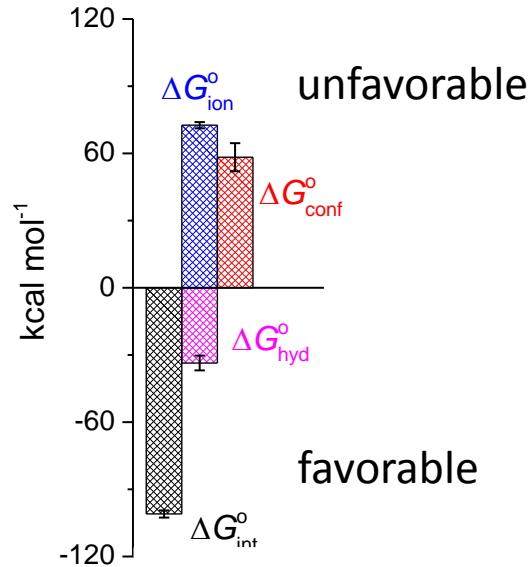


Thermodynamic driving forces

T = 25 °C



$$\Delta C_p^\circ = -400 \text{ cal mol}^{-1} \text{ K}^{-1}$$



M. Bončina et al. *Biophys. J.* 2015, in press.

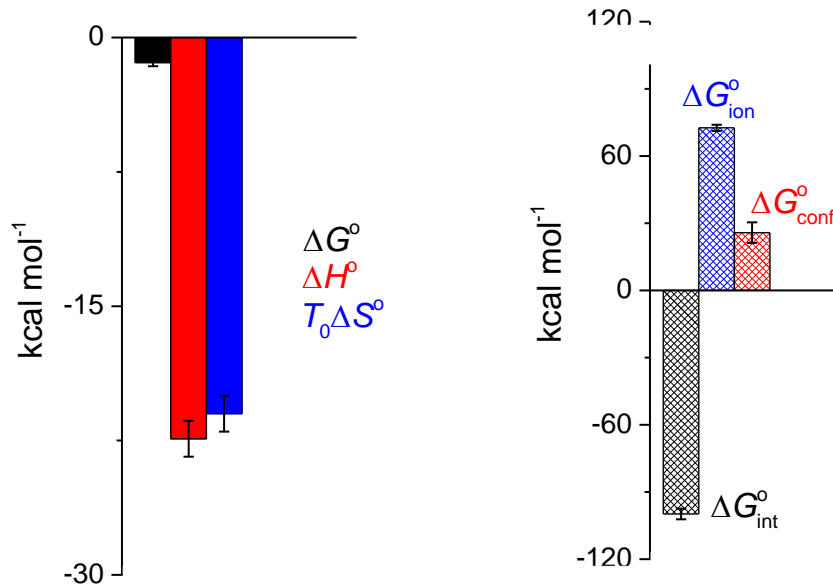
M. Bončina et al. *J. Am. Chem. Soc.* 2012, **134**, 9657.

H. Sugiyama et al. *J. Am. Chem. Soc.* 2010, **132**, 14910.

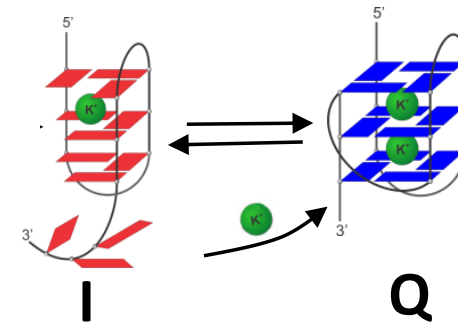
- Removal of hydrophobic groups (thymines) from water [$\Delta G^\circ_{\text{hyd}} < 0$].
- Specific interactions [$\Delta G^\circ_{\text{int}} < 0$].

Thermodynamic driving forces

T = 25 °C



$$\Delta C_p^\circ = 0 \text{ cal mol}^{-1} \text{ K}^{-1}$$



M. Bončina et al. *Biophys. J.* 2015, in press.

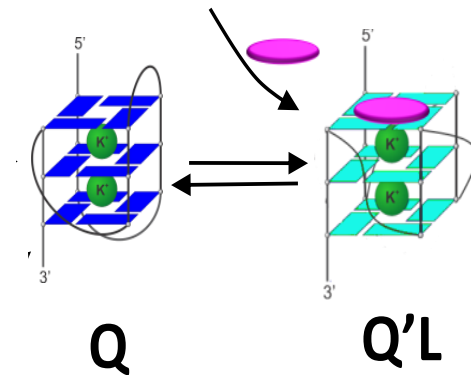
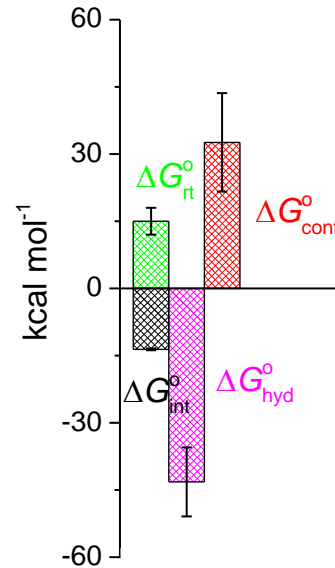
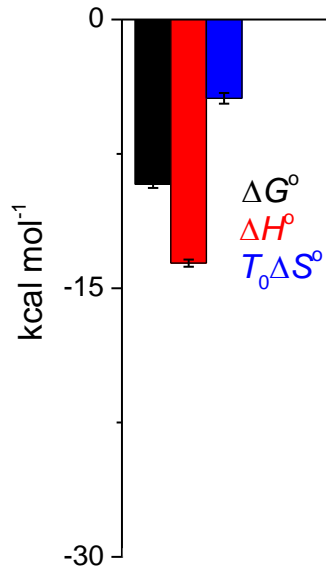
M. Bončina et al. *J. Am. Chem. Soc.* 2012, **134**, 9657.

H. Sugiyama et al. *J. Am. Chem. Soc.* 2010, **132**, 14910.

- Specific interactions [$\Delta G^\circ_{\text{int}} < 0$; $\Delta G^\circ_{\text{hyd}} \approx 0$].
- $\Delta G^\circ_{\text{conf}}(\text{U} \rightarrow \text{I}) \approx 2\Delta G^\circ_{\text{conf}}(\text{I} \rightarrow \text{Q})$ is in accordance with structural properties of I that are closer to Q than to U.

Thermodynamic driving forces

T = 25 °C

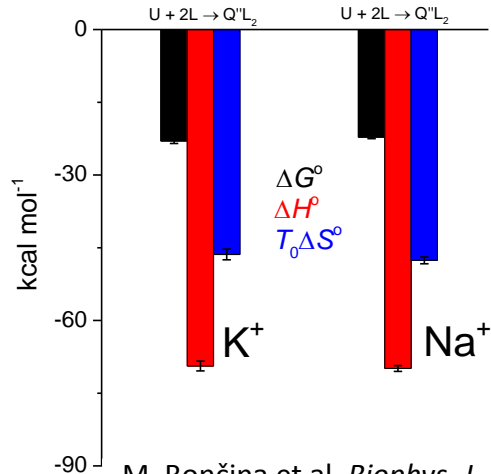


$$\Delta C_p^\circ = -540 \text{ cal mol}^{-1} \text{ K}^{-1}$$

M. Bončina et al. *Biophys. J.* 2015, in press.

- Removal of water from ligand-Tel22 interacting surface [$\Delta G^\circ_{hyd} < 0$].
- Specific (π - π stacking) interactions [$\Delta G^\circ_{int} < 0$] required for successful binding.
- Significant DNA conformational entropy loss [$\Delta G^\circ_{conf} > 0$].

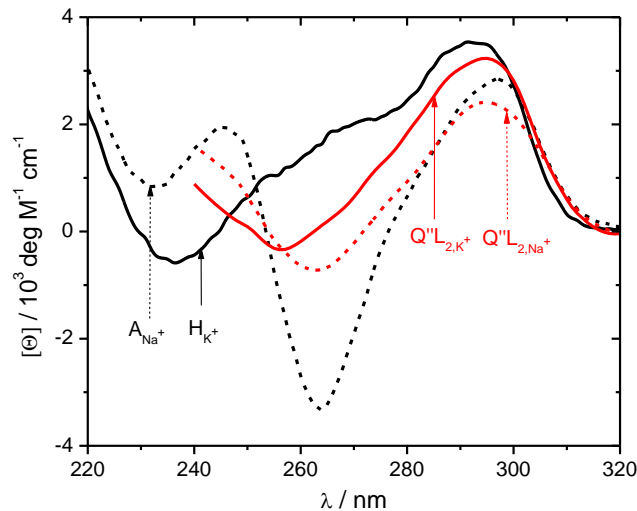
K^+ versus Na^+



M. Bončina et al. *Biophys. J.*, 2015, in press.

Thermodynamic analysis

The overall thermodynamics of binding-coupled folding is in the K^+ and Na^+ environment nearly the same.



CD spectroscopy

$Q''L_2(K^+) \approx Q''L_2(Na^+)$ but different to ligand-free H (K^+) or A (Na^+).

Meaningful findings

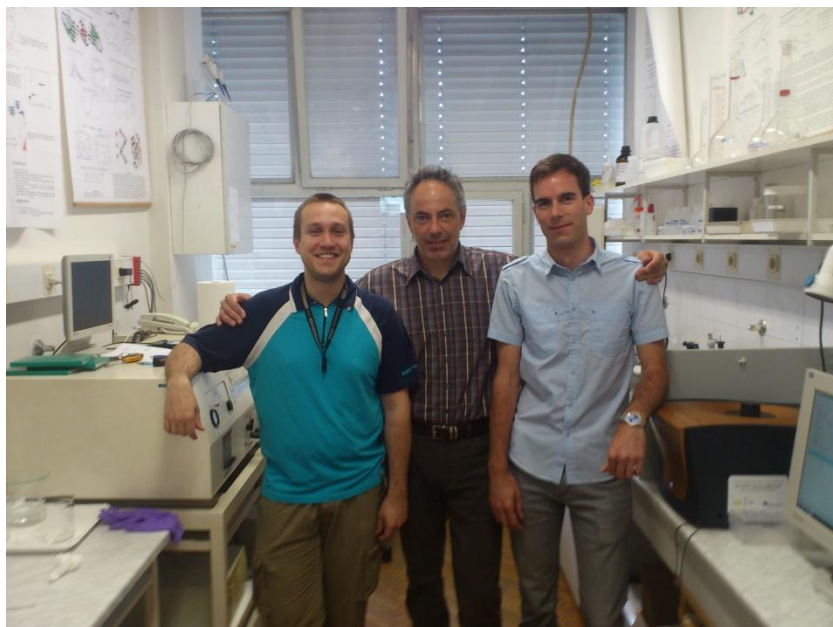
- Global thermodynamic analysis predicts:
 - thermodynamic forces that govern the formation of stable quadruplexes, folding intermediates and ligand-quadruplex complexes.
 - behavior of DNA fragments in the solution as a function of temperature and concentrations of solutes.

- Predictions are consistent with the observed structural features.

Thanks

Ljubljana (thermodynamics)

- Matjaž Bončina
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- Gorazd Vesnaver
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London (molecular modeling)

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